Supplementary materials

Optically Stimulated Luminescence (OSL) methodology

Sample preparation and analytical facilities

Luminescence samples were prepared under subdued red light condition in the luminescence laboratory from Cluj-Napoca, Romania. The material from the ends of each sample tube was used for gamma spectrometry and water content measurements whereas polymineral fine grains (4-11 µm) and coarse (63-90 µm) K-feldspars were extracted from the inner part. Samples were first treated with hydrochloric acid (10% concentration) for calcium carbonate removal, followed by a treatment with hydrogen peroxide (30% concentration) in order to remove the organic matter. All samples were wet sieved in order to collect the finer (<63 µm) and coarser (>63 µm) grains. Polymineral fine grains (4-11 µm) were obtained by settling using Stoke's law and centrifuging in distilled water (Frechen et al., 1996; Lang et al., 1996). Coarser grains in the range of 63-90 µm was obtained through dry sieving. For coarse K-rich feldspars extraction, an etching with HF (10% concentration) for 40 min was employed to remove the coating and alpha irradiated layer, and the material was then rinsed for 40 min with HCl (10% concentration) to remove any precipitated fluorides. K-feldspar enriched extracts were then separated using a heavy liquid solution with $\rho < 2.58$ g/cm^3.

Luminescence analyses were performed using TL/OSL Risø DA-20 reader equipped with automated detection and stimulation head (DASH) (Lapp et al., 2015). Luminescence signals were detected by PDM 9107Q-AP-TTL-03 (160-630 nm) photomultiplier. A blue filter combination (Schott BG36+Corning 7-59, with transmission between 320-460 nm) was used for the detection of polymineral fine grains and K-feldspar signals. Laboratory irradiations were carried out using a calibrated ^{90}Sr/^{90}Y beta source delivering 0.0764 Gy/s for fine grains and 0.086 Gy/s for coarse grains. K-feldspar coarse grains were mounted on stainless steel discs using silicone oil as an adhesive while for polymineral fine grains aluminium discs were used.

Equivalent dose determination

Equivalent doses on polymineral fine grains and K-feldspar coarse grains were determined using post-IR IRSL protocols based on SAR procedure with two different preheat and post-IR IRSL stimulation temperature combination (pIRIR$_{290}$ and pIRIR$_{225}$). For equivalent dose determination using pIRIR$_{290}$ protocol (see Table S1a) (Buylaert et al., 2011a; Buylaert et al., 2012; Buylaert et al., 2011b; Thiel et al., 2011) samples were preheated for 60 s at 320 °C and stimulated with IR diodes for 200 s at 50 °C to reduce the signal that is prone to fading by allowing recombination of near-neighbour trap and centre pairs. A subsequent IR stimulation at 290 °C for 200 s was employed in order to record the signal of interest. A high-temperature bleach was performed for 100 s at 325 °C at the end of each cycle.

Table S1: Flowchart of the pIRIR$_{290}$ (Buylaert et al., 2011a; Buylaert et al., 2012; Buylaert et al., 2011b; Thiel et al., 2011) and pIRIR$_{225}$ (Buylaert et al., 2009; Vasiliniuc et al., 2012) protocols applied in this study.

step	a. pIRIR$_{290}$	b. pIRIR$_{225}$
1	Dose	Dose
2	Preheat (320˚C; 60s)	Preheat (250˚C; 60s)
3	IRSL (50˚C; 200s)	IRSL (50˚C; 200s)
4	IRSL (290˚C; 200s)	IRSL (225˚C; 200s)
5	Test dose	Test dose
6	Preheat (320˚C; 60s)	Preheat (250˚C; 60s)
7	IRSL (50˚C; 200s)	IRSL (50˚C; 200s)
8	IRSL (290˚C; 200s)	IRSL (225˚C; 200s)
9	IRSL (325˚C; 100s)	IRSL (290˚C; 100s)

The second protocol used for equivalent dose determination was pIRIR$_{225}$ protocol (see Table S1b) (Buylaert et al., 2009; Vasiliniuc et al., 2012), which involves a preheat of 250 °C for 60s and an IR stimulation, firstly at 50 °C for 200s and then at 225 °C for 200s. The test dose response was determined in the same way. In order to reduce the residual charge, a bleaching at 290 °C for 100s was performed at the end of each cycle.

The size of the test dose used for equivalent dose determination varied between 20% and 50% of the equivalent dose value.

Annual dose determination

The radionuclide activity concentration was determined using high-resolution gamma spectrometry with a well-HPGe detector. The samples were stored for one month in order to achieve equilibrium between [226]Ra and [222]Rn. The conversion factors tabulated by Guérin et al. (2011) were used in determining the annual dose rates. For polymineral fine grains, an alpha efficiency factor of 0.08±0.02 was assumed. The water content was assumed to be 15% with a relative error of 25%. The total dose rates include the external contribution from beta and gamma radiation (additionally alpha radiation for 4-11 µm), as well as from the cosmic rays. The dose rates of cosmic rays were determined as a function of depth, altitude and geomagnetic latitude (Prescott and Hutton, 1994). An internal beta dose was included in the dose rate for K-feldspars, calculated assuming a K-content of 12.5±0.5%. For polymineral fine grains (4-11 µm) it was assumed that any dose rate derived from internal alpha activity is negligibly small. The radionuclide activity concentration and the annual doses for both samples are presented in Table S2.

Table S2: Summary of the pIRIR$_{225}$ and pIRIR$_{290}$ ages obtained on polymineral fine grains (4-11 μm) and coarse K-feldspars (63-90 μm) and dosimetry data. The ages were determined considering 15% water content.

Sample code	ED-pIRIR225		ED-pIRIR290		Gamma Spectrometry			Annual dose		Age (ka)-pIRIR225		Age (ka)-pIRIR290	
	pfg	K-feld. (63-90 μm)	pfg	K-feld. (63-90 μm)	Th-232	Ra-226	K-40	pfg	K-feld. (63-90 μm)	pfg	K-feld. (63-90 μm)	pfg	K-feld. (63-90 μm)
NZ 13A	55.5 ±1.9	60.6 ±1.6	72.7 ±3.3	78.8 ±3.3	29.3 ±0.9	32.9 ±0.7	530 ±17	3.48± 0.06	3.01± 0.04	16.0±1.4	20.1±1.5	20.9±2	26.2±2.1
NZ 14A	16.1 ±0.5	5.6± 0.2	20.9 ±1.7	9.5± 0.7	29.3 ±1.1	31.5 ±1.0	518 ±14	3.48± 0.06	3.03± 0.04	4.6±0.4	1.9±0.1	6.0±0.7	3.1±0.3

Fading test

It is well known that the infrared stimulated luminescence (IRSL) signal from feldspars is prone to fading (Wintle, 1973). Anomalous fading is a decrease of the IRSL signal with time, being a result of quantum mechanical tunnelling (Visocekas, 1985). Thus, the IRSL ages tend to underestimate the depositional age. In order to check if the IRSL signals from NZ13A and NZ14A samples were stable in time, 3 aliquots for each sample were used for the fading rates (percentage of the signal lost per decade of time (Aitken, 1985)) measurements using pIRIR$_{225}$ protocol. The aliquots used for g-value measurements were first utilized in residual dose determination. Each aliquot was irradiated with a beta dose of 50 Gy. A test dose of 25 Gy was used throughout all fading measurements. Prior to fading measurements, four consecutive prompt readouts were carried out. The IRSL signals of interest were then read-out after a preheat treatment and different storage times. After the longest storage time of 48 hours, two consecutive readouts were carried out.

Luminescence properties

Representative laboratory growth curves constructed for polymineral fine (Fig. S1a, c) grains as well as for coarse K-feldspars (Fig. S1b,d) from sample NZ13A using pIRIR$_{225}$ and pIRIR$_{290}$ protocols are shown in Fig. S1. The dose response curves were well fitted using a sum of two saturating exponential functions. For all aliquots measured, the recycling ratios were within 10% from unit indicating that the test dose signal successfully corrects for sensitivity changes during the repeated SAR cycles. As can be seen in the insets of Fig. S1, the decay of the natural and a regenerative signal have similar patterns. The recuperation ratios are small (<5%) for all aliquot measured.
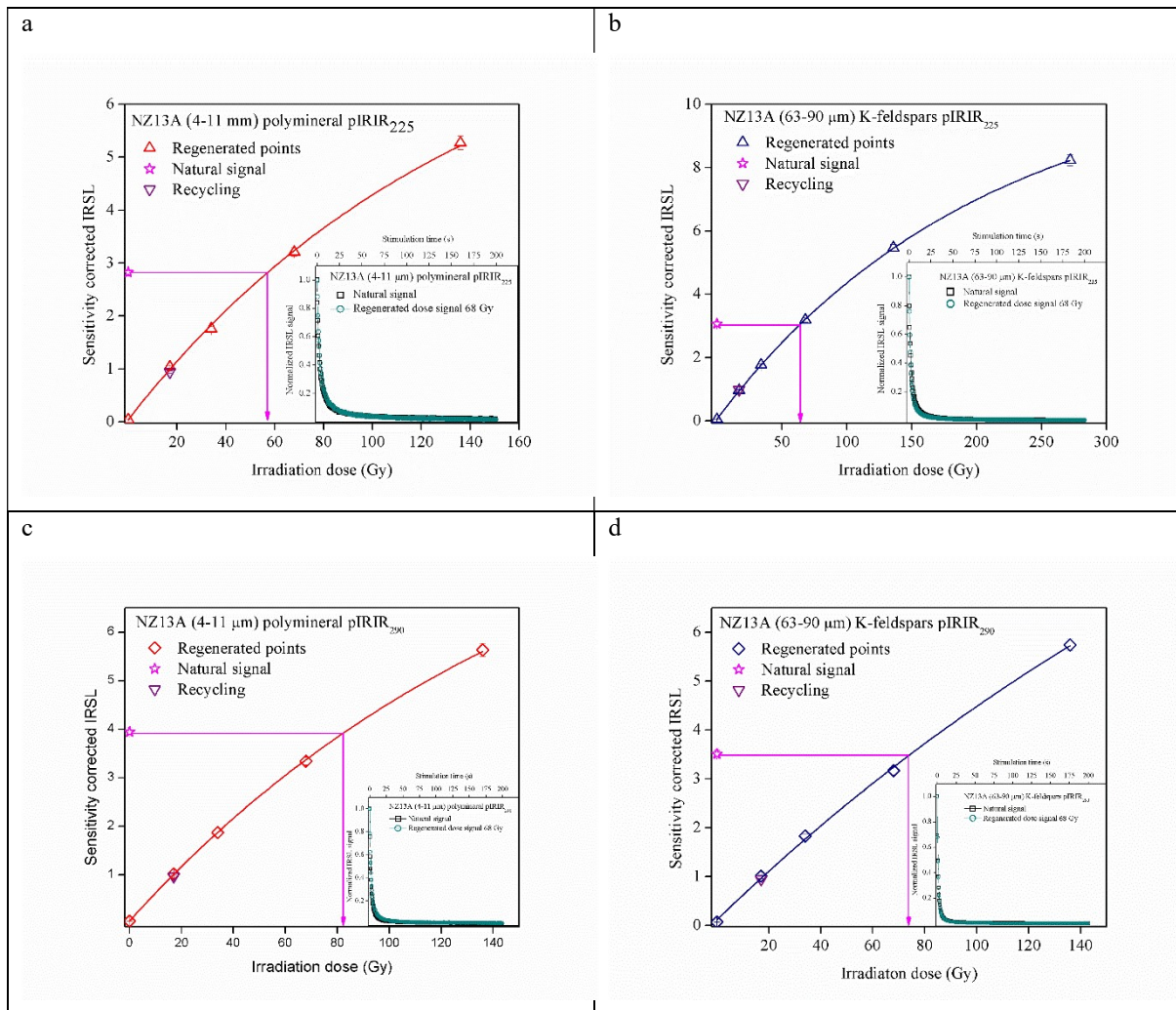
Figure S1: Representative sensitivity-corrected dose response curves constructed for sample NZ13A using one aliquot of (a) polymineral fine grains, (b) coarse K-feldspars using pIRIR₂₂₅ protocol and (c) polymineral fine grains, (d) coarse K-feldspars using pIRIR₂₉₀ protocol. The sensitivity corrected natural signal is represented as a star and the arrow indicates the equivalent dose. Recycling points are presented as an inverse triangle. The inset show a typical decay curve of natural IRSL signal (open square) in comparison to a regenerated signal (open circles) induced by a beta dose approximately equal with the equivalent dose.

Residual doses

Previous studies in literature have reported difficulties in bleaching the natural pIRIR signals even after prolonged exposure in daylight or in a solar simulator, obtaining residual doses of a few Gy (Buylaert et al., 2011b; Lowick et al., 2012; Roberts, 2012).

In order to determine the residual doses, three to five fresh aliquots were exposed for 20 days to daylight. The residual doses obtained for polymineral fine grains are 3.3±0.3 Gy (NZ13A) and 3.0±0.2 Gy (NZ14A) using pIRIR₂₂₅ protocol and 10±1 Gy (NZ13A) and 11±1 Gy (NZ14A) using pIRIR₂₉₀ protocol. In the case of coarse K-feldspars residual doses of 2.9±0.1 Gy (NZ13A) and 1.3±0.1 Gy (NZ14A) was obtained using pIRIR₂₂₅ protocol

and 9.0±0.3 Gy (NZ13A) and 4.9±0.3 Gy (NZ14A) using pIRIR$_{290}$ protocol. Similar values for the residual doses have been reported so far in literature. Murray et al. (2014) reported pIRIR$_{290}$ residual doses ranging between 4.5±0.3 and 10.1±0.1 Gy for loess samples collected from Serbia, SE Europe and Sohbati et al. (2016) measured an average residual dose of 6.15±0.10 Gy using pIRIR$_{290}$ protocol on loess samples from New Zealand, South Island. Also, low residual doses were obtained using pIRIR$_{225}$ protocol by Buylaert et al. (2009) for samples with various sedimentary origins.

Dose recovery

The most stringent test for any SAR protocol is the dose recovery test (Murray, 1996; Murray and Wintle, 2003; Wallinga et al., 2000), that analyses whether the protocol can successfully measure a known irradiation dose prior to any thermal treatment.

A dose recovery test was performed on five aliquots for polymineral fine grains and five aliquots on coarse K-feldspars for each sample (NZ13A and NZ14A) using both protocols (pIRIR$_{225}$ and pIRIR$_{290}$). The fresh aliquots were exposed for 21 days to daylight in order to remove the natural signals. Laboratory known doses were chosen to be approximately equal to the measured equivalent dose. The dose recovery ratios were calculated after the residual doses were subtracted from the measured dose. The size of the test dose used for dose recovery measurements varied between 20-50% of the size of the given dose. Figure S2 shows the results for the dose recovery test for sample NZ13A (Fig. S2a) and NZ14A (Fig. S2b). The ratios between the recovered and given dose for polymineral fine grains are 1.11±0.03 (NZ13A) and 1.01±0.03 (NZ14A) using piRIR$_{225}$ protocol and 1.14±0.05 (NZ13A) and 1.03±0.06 (NZ14A) using pIRIR$_{290}$ protocol. In the case of coarse K-feldspars, the dose recovery ratios are 0.96±0.01 (NZ13A) and 0.87±0.03 (NZ14A) using pIRIR$_{225}$ protocol, while those obtained by applying pIRIR$_{290}$ protocol are 1.17±0.06 (NZ13A) and 1.16±0.04 (NZ14A). We conclude that the overall behaviour of the pIRIR$_{225}$ protocol is satisfactory whereas the pIRIR$_{290}$ protocol overestimates the given dose by ~ 17%.
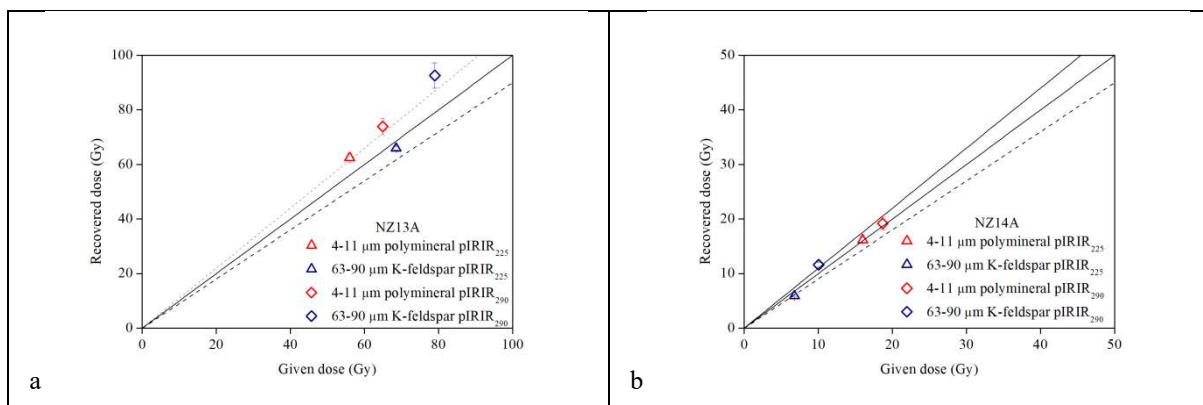
Figure S2: Dose recovery test results for polymineral fine grains and coarse K-feldspars using pIRIR$_{225}$ and pIRIR$_{290}$ protocols for (a) NZ13A and (b) NZ14A samples. The given doses were chosen to be approximately equal to the equivalent dose for each sample. The solid line indicates the ideal 1:1 dose recovery ratio while the dash lines represent a 10% variation from unity.

Equivalent doses

The equivalent doses measured on polymineral fine grains as well as on coarse K-feldspars grains using pIRIR$_{225}$ and pIRIR$_{290}$ protocols are displayed in Table 2. For NZ13A sample an equivalent dose of 55.5±1.9 Gy and 60.6±6 Gy was measured using pIRIR$_{225}$ protocol on polymineral fine grains and coarse K-feldspars, respectively. For the same sample, the equivalent dose using pIRIR$_{290}$ protocol is 72.7±3.3 Gy on polymineral fine grains while on coarse K-feldspars a value of 78.8±3.3 Gy was obtained. In the case of NZ14A sample, the equivalent doses obtained using pIRIR$_{225}$ protocol are 16.1±0.5 Gy on polymineral fine grains and 5.6±0.2 Gy on coarse K-feldspars, whereas using pIRIR$_{290}$ protocol are 20.9±1.7 Gy on polymineral fine grains and 9.5±0.7 Gy on coarse K-feldspar. The equivalent doses presented in Table 2.

g-values

g-values were measured for both samples (NZ13A and NZ14A) using pIRIR$_{225}$ protocol. Negative g-values were obtained for polymineral fine grains in the case of these two samples. For coarse K-feldspars an average g-value for sample NZ13A was 0.60±0.5%/decade, while for sample NZ14A the measured value was that of 0.85±0.09%/decade. Figure S3 shows the results obtained for the fading measurement for a representative aliquot from sample NZ13A for polymineral fine grains (Fig. S3a) and coarse K-feldspars (Fig. S3b). As it can be seen in Fig. S3, the measured sensitivity corrected IRSL signals after different delay time's scatter in the range of the signals obtained after the 6 prompt readouts. We conclude that such low fading rates represent laboratory artefact and do not accurately reflect the signal stability in nature (Buylaert et al., 2012; Roberts, 2012; Thiel et al., 2011). Therefore the ages presented in this study were the ones not corrected for fading.
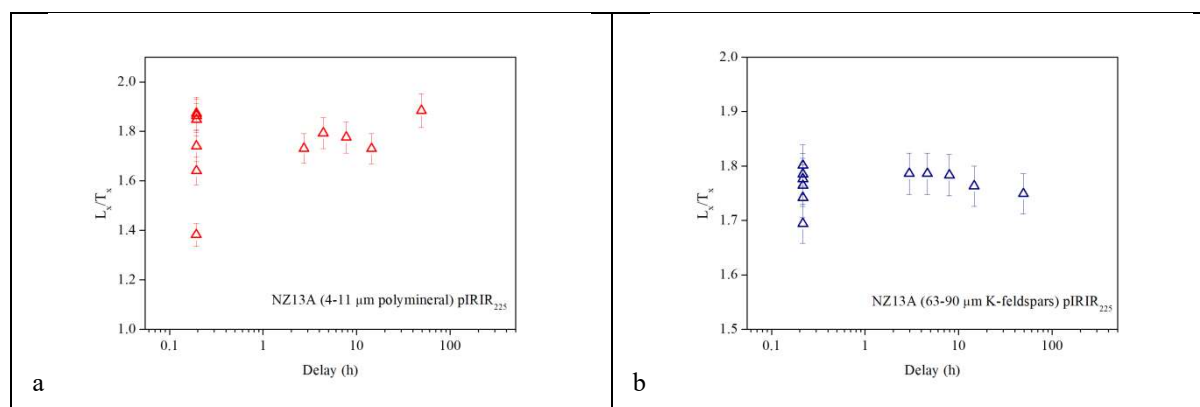
Figure S3: Results of the fading rates measurement using pIRIR$_{225}$ protocol on one representative aliquot from sample NZ13A for (a) polymineral fine grains and (b) coarse K-feldspars. A maximum delay of 48 hours was used for both samples. A number of 4 consecutive prompt readouts were carried out before signal measurement after delay and two consecutive prompt readouts were added after the 48 hours delay.

Various values of the fading rates were reported in literature for the IRSL signal measured using pIRIR$_{225}$. g-values >1 %/decade were obtained (Buylaert et al., 2009; Thomsen et al., 2008; Zhang et al., 2018), while Vasiliniuc et al. (2012) calculated g-values ranging from 0.6 to 1.3 %/decade. Previous studies reported low fading

rates (<1%/decade) measured by using pIRIR$_{290}$ protocol. Sohbati et al. (2016) measured g-values of 0.9±0.3 %/decade for New Zealand loess, while Thiel et al. (2011) reported g-values considered to be laboratory artefact for Austrian loess.

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